

Konakhin, a Novel Type of Degraded Sesterterpene; Isolation from a Marine Sponge of Senegal

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An unidentified sponge collected off Konakhè, near Dakar, contains a novel type of degraded linear sesterterpene, konakhin (+)-**1**, formally deriving from the replacement by chlorine of the carbonyl carbon in the tetronic moiety of fasciculatin (–)-**2**, the main terpene of this sponge.

Sesterterpenes¹ have been rare natural products until the study of the marine sponges of the order Dictyoceratida and the nudibranch molluscs, that feed on them,² which have given a wide variety of compounds of this class and degraded derivatives.³

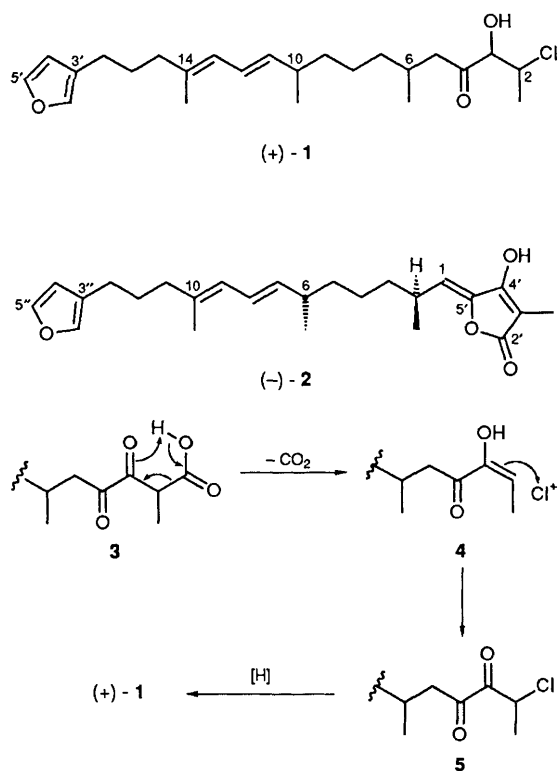
We report here on a novel type of degraded sesterterpene isolated from an undetermined marine sponge collected at –30 m off Konakhè, near Dakar. The compound, konakhin (+)-**1**,[†] is a colourless oil, $[\alpha]_D^{20} + 6.0$ (*c* 0.1, CHCl₃) whose molecular formula C₂₄H₃₇ClO₃ was derived from ¹³C NMR and mass spectra.[‡] The partial structure from the furanic ring

up to C-7 rests on the superimposition of ¹³C and ¹H NMR signals with the corresponding signals of fasciculatin (–)-**2**; this is the main terpene of our sponge, already isolated from the Mediterranean sponges *Ircinia fasciculata*⁴ and *Ircinia variabilis*⁵ and assigned the Z-1 configuration.⁶ The remaining portion of (–)-**1** is supported by NMR spectra, which reveal keto, CHOH, and CHCl groups from the characteristic signals at δ 207.96 s, 84.22 d and 55.39 d, respectively. That the latter two groups represent C-3-H(OH) and C-2-HCl stems from the heterocorrelation of the δ(C) 84.22 d with the δ(H) 4.53 d and the δ(C) 55.39 d with the δ(H) 4.29 dq. The C-6-H-C-5-H₂ group is supported by an ABX pattern where the δ(H) values 2.83 and 2.57 and *J*_{AB} 19 are consistent with a keto group at C-4; changing to CDCl₃ as solvent, the OH group becomes a br, s at δ(H) 2.22, indicating an intramolecular hydrogen bond

[†] *Ca.* 0.006% on dry sponge weight after ethanol extraction, evaporation, flash chromatography on Merck Kieselgel 60 with *n*-hexane–AcOEt and acetylation of the most polar fractions with Ac₂O–pyridine. (MeOH) λ_{max}/nm 238 (ε 22 000); IR (neat) ν_{max}/cm^{–1} 3480s, 1720m.

[‡] ¹³C NMR spectrum (CDCl₃, 75.43 MHz, H-bearing C assigned by HETCOR) δ 20.96 (q, C-1), 55.39 (d, C-2), 84.22 (d, C-3), 207.96 (s, C-4), 48.38 (t, C-5), 27.95 (d, C-6) 36.85 (t, C-7), 24.75 (t, C-8), 37.34 (t, C-9), 37.01 (d, C-10), 138.49 (d, C-11), 124.70 (d, C-12), 125.11 (d, C-13), 135.97 (s, C-14), 39.34 (t, C-15), 28.11 (t, C-16), 24.38 (t, C-17), 20.63 (q, *Me*-C-6), 19.51 (q, *Me*-C-10), 16.51 (q, *Me*-C-14), 138.77 (d, C-2'), 124.98 (s, C-3'), 110.97 (d, C-4'), 142.63 (d, C-5'). EIMS (*m/z*, %) 410/408 (1/3 M⁺), 395/393 (7/21, M⁺ – Me), 372 (14, M⁺ – HCl), 357 (57, M⁺ – Me – HCl), 135 (100). High resolution EIMS *m/z* 408.2410 (C₂₄H₃₇ClO₃ requires 408.2431).

§ ¹H NMR spectrum (CD₃OD, 299.94 MHz, *J* in Hz) δ 1.57 (d, *J*_{1,2} 6.8, 3H-C-1), 4.29 (dq, *J*_{2,1} 6.8, *J*_{2,3} 5.6, H-C-2), 4.53 (d, *J*_{3,2} 5.6, H-C-3), 2.83 and 2.57 (AB part of ABX, *J*_{AB} 18.8, *J*_{AX} 5.1, *J*_{BX} 7.8, 2H-C-5), 2.00 (X part of ABX, m, H-C-6), 1.24 (series of m, 2H-C-7, 2H-C-8, and 2H-C-9), 2.15 (m, H-C-10), 5.40 (dd, *J*_{11,12} 16.0, *J*_{11,10} 8.3, H-C-11), 6.21 (ddd, *J*_{12,11} 15.0, *J*_{12,13} 10.7, *J*_{12,10} 0.8, H-C-12), 5.77 (br, d, *J*_{13,12} 10.7, H-C-13), 2.07 (t, *J*_{15,16} 7.9, 2H-C-15), 1.68 (pseudo quint., *J* 7.8, 2H-C-16), 2.39 (t, *J*_{17,16} 7.6, 2H-C-17), 7.26 (m, H-C-2'), 6.30 (m, H-C-4'), 7.37 (pseudo t, *J* 1.8, H-C-5'), 0.88 (d, *J*_{Me,6} 6.6, *Me*-C-6), 0.99 (d, *J*_{Me,10} 6.6, *Me*-C-10), 1.73 (d, *J*_{Me,13} 1.2, *Me*-C-14), (in CDCl₃ δ 2.22 br.s, OH).



Scheme 1

with the adjacent carbonyl group; the latter point is further supported by the fact that (+)-1 resists acetylation in AC_2 -pyridine at room temp.

A biogenetic route from fasciculatin to konakhin can be envisaged as in Scheme 1. Thus, decarboxylation of fasciculatin in the carboxylic form **3** may be viewed to give the enol **4** which undergoes chlorination by a haloperoxidase-provided chloronium ion² to afford the diketone **5** which is finally

reduced to konakhin (+)-1. In any event, the degradation implied by the structure of konakhin (-)-1 has no precedent and, in general, halogenated degraded terpenes are rare compounds, an example being the trinorsesquiterpene kumepaloxane, isolated from a nudibranch.⁷

In the hope to shed further light on the behaviour of tetrone acid terpenes, we are trying the difficult isolation of other unusual terpenes present in trace amounts in our sponge while carrying out degradation studies of these and model compounds; we hope to present the entire study in a full paper.

We thank Mr A. Sterni and L. Zuppiroli and for MS measurements and M.P.I. (Progetti di Interesse Nazionale and Project for the Cooperation between the Università di Trento and the Université Cheikh Anta Diop) and C.N.R., Roma, for financial support.

Received, 31st July 1990; Com. 0/03503C

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